# MICROORGANISMS CAUSING DETERIORATION OF STORED GARLIC

## Aruni Chantarasnit\* and Vimol Phanichyakarn\*\*

#### ABSTRACT

Samples of garlic in storage sheds in three northern Thai provinces were examined for fungi and other organisms causing deterioration. Twenty-three genera of fungi were found in 71 samples tested. Aspergillus spp. were found in nearly all samples. Other common fungi were Penicillium and Curvularia. Three field pathogens, namely Fusarium, Colletotrichum and Alternaria, were also found. Apparently normal looking cloves were often found to be contaminated inside. Bacterial infection was commonly found and resulted in rot. Rates of contamination by microorganisms were significantly reduced by surface desinfection. Several animal pests including insect larvae and mites occurred on the same deteriorating garlic.

## INTRODUCTION

Garlic (Allium sativum Linn.). a plant in the family Liliaceae, has been useful to mankind since ancient times. The Chinese people use the plant as a vegetable. They and people in other countries have admired garlic as a spice or herb for improving one's health, despite its strong odor, and its therapeutic value has recently been recognized (IKRAM, 1972; JAIN, 1977; BHUSHAN et al., 1979; BOULLIN, 1981). Therefore, the present consumption of garlic is increasing.

Garlic is economically important as a second crop after rice in northern Thailand. The bulbs are harvested in March and stored for use during the year (Fig.1). Deterioration occurs gradually during this period, reducing the quality as well as the quantity of the product. In certain years, the price soars a few months before the new crop is harvested due to the low supply.

In spite of the inhibitory effect of the allicin substance in garlic extract (CAVALLITO & BAILEY, 1944; KABELIK, 1970; IMWIDTHAYA et al., 1978), many microorganisms including fungi grow on stored garlic bulbs and thus damage the product (SMALLEY & HANSEN, 1962; GEOGIEVA & KOTEV, 1979). A number of Aspergilli and Penicillia have been reported to produce various kinds of aflatoxins on certain kinds of substrate (SHANK et al., 1972). Therefore, the objectives of our study were as follows:

- 1. To investigate the growth of microorganisms found on garlic from the North and to evaluate their significance in destroying stored garlic and/or spreading diseases if infected cloves are seeded.
- 2. To isolate fungi which are suspected of producing mycotoxin for further studies.

<sup>\*</sup> Department of Botany, Faculty of Science, Chulalongkorn University, Bangkok 10500, Thailand.

<sup>\*\*</sup> Department of Biology, Faculty of Science, Chulalongkorn University, Bangkok 10500, Thailand.

## METHODS

Samples of garlic bulbs were collected from six storages in the northern provinces of Chiang Mai, Lamphun and Lampang. Each month during the 1-year study period (July 1983 – June 1984), a sample of 1 kg was taken from each selected storage shed. A subsample of 100 g was taken from each 1-kg sample and was subdivided into two equal parts of 50 g. Each bulb in the first part was split into cloves and pretreated with 20% chlorox solution before being incubated on sterile moist blotters (Fig.2). The second part was incubated similarly without pretreatment. Samples were examined for microorganisms after 1 week of incubation.

The percentage of decayed cloves and percentage of cloves infected by each type of microorganism in each sample were recorded.

## RESULTS

Fungi Encountered. Twenty-three genera of fungi were observed from 71 tested samples (Table 1). Aspergillus was most frequently found followed by Penicillium, Curvularia and Fusarium; the numbers of samples contaminated by these fungi were 70, 64, 33 and 29 respectively.

The Most Destructive Species. A. niger was found in almost all of the tested samples and in a much higher percentage than any other species. It thrived on the surface as well as inside the cloves. Normal looking bulbs sometimes contained fungal spoilage inside. Its growth characteristics on garlic and those of A. flavus, Penicillium sp., Curvularia sp., and Fusarium sp. are shown in Table 2 and Fig.3. Sometimes two or more types of fungi were present on the same clove (Fig.4). Bacterial infection associated with insect damage can also result in high percentage of garlic rot (Fig.5).

Pretreatment with Diluted Chlorox Solution. Samples receiving pretreatment of 20% chlorox solution showed a lower percentage of fungal and bacterial contamination (Table 3 and Fig. 6). This indicates that more microorganisms exist on the surface of the cloves than inside. Invasion into the cloves usually occurred through wounds made by insects, or by mite transmission.

Occurrence of Field Pathogens. Alternaria sp., Colletotrichum sp., and Fusarium sp., reported as causing garlic diseases in the field (BRANDAO & SOARES, 1943; GIATIGONG, 1980), were also encountered in our samples. The first two fungi occurred on a few samples, while Fusarium spp. were more common, especially from samples harvested from the infected field. Immature garlic showed a higher percentage of infection than did mature garlic. Newly harvested samples also gave higher percentages of Fusarium infection, which was gradually replaced by that of the storage fungi (Aspergillus spp.) during the time of storage (Table 4). Therefore, the chance of being the first inoculum in the field for the following planting season is low.

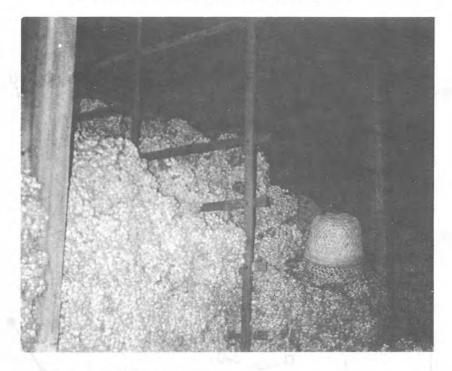


Figure 1. Garlic in storage.

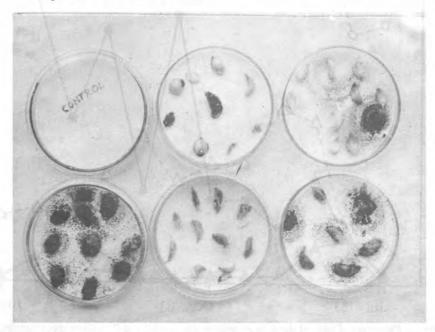


Figure 2. Incubation of tested garlic samples on sterile moist blotters.

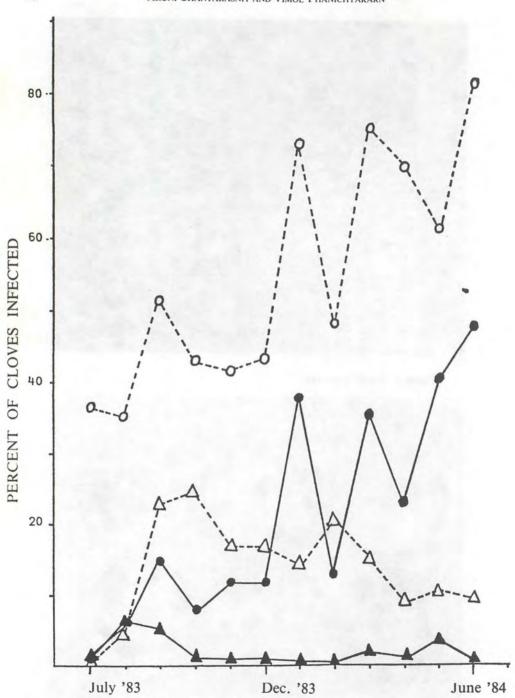


Figure 3. Percentage of infected cloves by *Aspergillus* spp. (circles) and *Penicillium* spp. (triangles) in each month during the study. Solid lines indicate surface disinfection treatment; dashed lines indicate no disinfection.

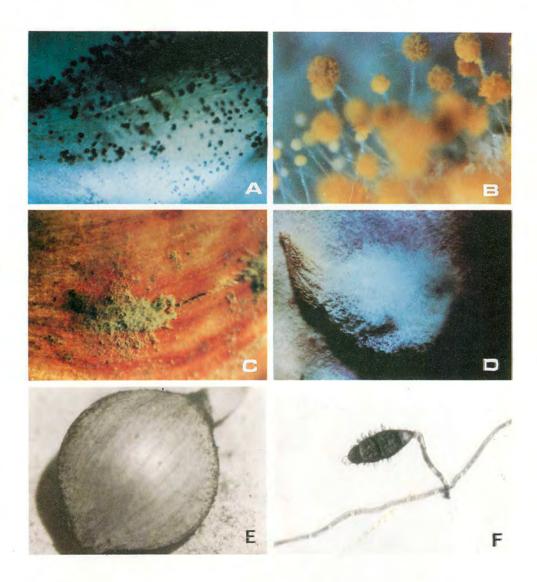


Figure 4. Growth of some fungi on garlic cloves. A, Aspergillus niger; B, A. flavus; C, Penicillium funiculosum; D, Fusarium moniliforme; E, Curvularia tuberculata and F, Conidium of C. tuberculata.

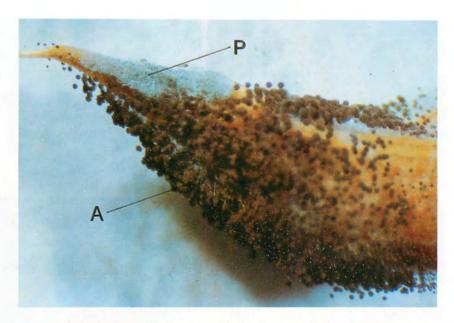


Figure 5. Aspergillus niger (A) and Penicillium sp. (P) on a garlic clove.



Figure 6. Bacterial infection resulting in garlic rot.

**Deterioration by Other Pests.** Besides fungi and bacteria, mites and insects also infested stored garlic. In the beginning of our investigation, spoilage of garlic due to mites was high, but was surpassed by spoilage due to microorganisms toward the end of the period of investigation.

#### ACKNOWLEDGEMENTS

The authors wish to thank Chulalongkorn University for granting funds for this study. We are deeply indebted to Associate Professor Suthasnee Boonkong who not only arranged the funds but also encouraged and helped us in various ways throughout the study. We are also grateful to Dr. W.Y. Brockelman and Miss Kerri Wright who kindly reviewed the manuscript, and to many individuals who assisted us during the survey of the stored garlic.

Table 1. Genera of fungi and percentages of infected garlic cloves.

	Genus	%	
1.	Alternaria	0.03	<del>.</del> ,
<i>2</i> .	Aspergillus	55.09	
<i>3</i> .	Chaetomium	0.48	
4.	Cladosporium	0.68	
5.	Colletotrichum	trace	
6.	Curvularia	1.24	
<i>7</i> .	Fusarium	1.17	
8.	Gliocladium	trace	
9.	Helminthosporium	trace	
<i>10</i> .	Mucor	trace	
11.	Neurospora	0.18	
<i>12</i> .	Nigrospora	0.04	
<i>13</i> .	Ovularia	trace	
14.	Penicillium	14.04	
<i>15</i> .	Pulluraria	0.06	
<i>16</i> .	Rhizoctonia	0.10	
<i>17</i> .	Rhizopus	0.47	
<i>18</i> .	Scopularia	0.01	
<i>19</i> .	Spicaria	trace	
<i>20</i> .	Syncephalastrum	0.05	
<i>21</i> .	Thamidium	trace	
<i>22</i> .	Trichoderma	0.50	
<i>23</i> .	Verticillium	trace	

Table 2. Growth characteristics and symptoms produced on garlic cloves by some fungi.

Fungi	Growth characteristics	Symptoms		
Aspergillus niger A. flavus	Scattered colonies on surface or occasionally under the clove's sheath	Dry rot and heart rot, discoloration		
Penicillium spp.	Dense colonies on surface	Dry rot, discoloration (P. funiculosum turned the sheath into pink)		
Fusarium moniliforme	Pinkish white colony, sometimes covering the whole clove	Discoloration and decay of immature garlic		
Curvularia tuberculata	Scanty mycelium, dark colonies with	Discoloration		
(C. lunata and C. palescens were more common)	dark conidia on short conidiophore			

Table 3. Percentages of infected cloves from each storage shed (averaged over 12 months).

	Without surface disinfection							With surface disinfection							
Organisms	1	2	3	4	5	6	Mean	1	2	3	4	5	6	Mean	L.S.D. for means
Aspergillus	65.3	60.0	40.3	44.3	43.0	73.6	54.42	26.6	25.5	20.8	16.7	17.2	28.2	22.55	8.21
Chaetomium	1.4	0.1	0.4	0.1	0.9	0.0	0.47	0.6	0.1	0.1	0.0	0.3	0.0	0.18	0.42
Curvularia	1.3	1.8	0.9	2.8	0.4	0.6	1.3	0.3	0.7	0.0	0.4	0.4	0.1	0.31	0.60
Fusarium	0.6	0.8	0.5	0.9	3.9	0.7	1.2	0.1	0.0	0.1	0.2	0.6	0.2	0.2	0.50
Penicillium	13.1	9.6	17.7	13.8	16.4	9.7	13.3	1.3	1.6	1.3	2.5	3.8	0.5	1.8	3.00
Trichoderma	1.4	0.7	0.6	0.3	0.0	0.4	0.6	3.5	0.7	0.02	1.6	4.5	0.3	1.7	1.00
Bacteria	14.5	17.4	9.8	15.4	20.4	13.9	15.2	3.4	7.7	4.7	8.7	10.1	7.2	6.9	4.02

Least Significant Difference (L.S.D.) between without and with surface disinfection for each storage:

Aspergillus	=	19.34	Penicillium	=	7.40
Curvularia	=	1.45	Trichoderma	=	2.35
Fusarium	=	1.07	Bacteria	=	9.97

Table 4. Percentage of cloves infected by Fusarium spp. and Aspergillus spp. (Samples of mature and immature garlic were taken from Fusarium-infected field).

Weeks after	Fusarium	infection	Aspergillus infection			
harvest	Mature clove	Immature clove	Mature clove	Immature clove		
2	13.0	44.0	0.0	1.0		
15	3.3	17.1	8.7	3.4		
36	0.0	4.1	15.7	22.3		

#### REFERENCES

BHUSHAN, S., S.P. SHAMA, S.P. SINGH, S. AGRAWAL, A. INDRAYON and P. SETH. 1979. Effect of garlic on normal blood cholesterol level. *Indian J. Physiol.* 23 (3): 211-214.

BOULLIN, D.J. 1981. Garlic as a platelet inhibitor. Lancet (8223), 766-777.

BRANDAO, F. and J. SOARES. 1943 Some diseases of garlic. Biol. Min. Agric. 30 (6): 26-34, 1941, in Biol. Abstr. No. 14751. Vol. 17

CAVALLITO, C.J. and J.H. BAILEY. 1944. Allicin, the antibacterial principle of Allium sativum.
I. Isolation, physical properties and antibacterial action. J. Amer. Chem. Soc. 66: 1950-1951.

GEOGIEVA, M. and S. KOTEV. 1979. The pathogens of garlic rot during storage (abstract). Review Plant Pathol. 83: 989.

Giatgong, P. 1980. Host Index of Plant Disease in Thailand. Mycology Branch, Plant Pathol. and Microbiol. Div., Dept. of Agric., Bangkok. pp. 2-3.

IKRAM, M. 1972. Review on chemical and medicinal aspects of Allium sativum. Park. J. Sci. Ind. Res. 15 (1-2): 81-86.

IMWIDTHAYA, S., A. PLANGPATANAPANICHYA and P. THASNAKORN. 1978. Antifungal activity of garlic extract. *Bull. Med. Tech. (Mahidol Univ.)*, 2(2): 68-78.

JAIN, R.C. 1977. Effect of garlic on serum lipids, coagulability and fibrinolytic activity of blood. Am J. Clin. Nutri. 30(9): 1380-1381.

KABELIK, J. 1970. Antimicrobial properties of garlic. *Pharmazie* 25(4): 266-270. In *Plant Biochem*. *Abstr.* 73: 83.

SHANK, R.C., G.N. Wogan and J.B. Gibson. 1972. Dietary aflatoxins and human liver cancer.

I. Toxigenic moulds in food and food stuff of Tropical South East Asia. Fd. Cosmet.

Toxicol. 10: 51-60.

SMALLEY, E.E. and H.N. HANSEN. 1962. Pencillium decay of garlic. Phytopathol. 52(7): 666-678.